

Journal Article

Calcium ion regulation of sodium alginate in pure buckwheat noodles shown by in vitro simulated digestion

Wang, H., Zhang, J., Han, L., Cao, J., Yang, J., Zhang, Y., and Hu, B.

This article is published by Frontiers Media. The definitive version of this article is available at: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9877419/>

Recommended citation:

Wang, H., Zhang, J., Han, L., Cao, J., Yang, J., Zhang, Y., and Hu, B. (2023) 'Calcium ion regulation of sodium alginate in pure buckwheat noodles shown by in vitro simulated digestion', *Frontiers in Nutrition*, 9: 1105878. doi: 10.3389/fnut.2022.1105878

Calcium ion regulation of sodium alginate in pure buckwheat noodles shown by *in vitro* simulated digestion

1 **Hongyan Wang¹, Jiukai Zhang³, Lingyu Han¹, Jijuan Cao^{1,2*}, Jixin Yang⁴, Ying Zhang¹, Bing**
2 **Hu^{1*}**

3 ¹College of Life Sciences, Key Laboratory of Biotechnology and Bioresources Utilization, Dalian
4 Minzu University, Ministry of Education, Dalian 116029, China;

5 ²Collaborative Innovation Center of Provincial and Ministerial Co-construction for Marine Food Deep
6 Processing, Dalian Polytechnic University, Dalian, 116034, China;

7 ³ Chinese Academy of Inspection and Quarantine, 11 Ronghua Nanlu, Yi Zhuang, Beijing 100176,
8 China

9 ⁴ Faculty of Arts, Science and Technology, Wrexham Glyndwr University, Plas Coch, Mold Road,
10 Wrexham, LL11 2AW, United Kingdom

11

12 * Correspondence:

13 Dr. Bing Hu; Tel: 86041187656215

14 E-mail address: hubing@dlnu.edu.cn

15 Dr. Jijuan Cao; Tel: 86041187656141

16 E-mail address: caojijuan@dlnu.edu.cn

17 **Keywords: buckwheat noodles, sodium alginate, calcium ions, *in vitro* simulated digestion,**
18 **release of glucose**

19 Abstract

20 The effects of calcium sodium alginate on quality and starch digestion of pure buckwheat noodles were
21 investigated. The incorporation of calcium ions into noodles containing sodium alginate was found to
22 reduce water absorption by the noodles during cooking, together with an increase of the turbidity.
23 Calcium addition improved the noodle texture, as shown by the measurement of hardness, elasticity,
24 adhesion, and chewability. *In vitro* simulations of digestion showed that calcium ion cross-linking with
25 sodium alginate reduced glucose formation by approximately 23.3 mg/g. X-ray diffraction and Fourier
26 transform infrared spectroscopy showed alterations in the crystal zone of the noodles induced by an
27 alginate gel network, although no new chemical substances were generated. Noodles prepared by this
28 exogenous method may be useful as functional foods for patients with diabetes.

29

30

31 1 Introduction

32 Starch is an essential carbohydrate component of the human diet. Hydrolysis by amylase reduces the
33 starch to glucose that can then be utilized by the body. However, starch, especially in the form of
34 processed foods, tends to be rapidly digested, leading to potential health issues such as obesity and
35 diabetes, and thus people are becoming more aware of the importance of consuming healthy food.(1,
36 2)

37 Noodles are a staple food in many Asian countries and have become increasingly popular among
38 consumers around the world.(3) However, noodles have a high glycemic index (GI) indicating that
39 they are easily hydrolyzed.(4) It is thus important to attempt to reduce the GI of noodles. Buckwheat
40 and its processed products contain a variety of bioactive substances, including specific polysaccharides,
41 dietary fiber, and polyphenols, all of which are of great nutritional values.(5) Therefore, buckwheat
42 noodles have attracted increased consumer attention.(6) It has been found that buckwheat noodles can
43 slow down sugar release resulting from starch digestion *in vitro* and that the addition of 45% Tartary
44 buckwheat to noodles can be beneficial to human health.(7)

45 It is well documented that adding functional components to starchy foods can slow the rate of
46 digestion.(8-10) Sodium alginate is a natural polysaccharide obtained from brown algae. It is a linear
47 compound composed of (1→4)-linked β -D-mannuronic acid and α -L-guluronic acid pyranose
48 residues.(11) Sodium alginate, being highly hydrophilic, has good hygroscopic properties and
49 dissolves readily in both hot and cold water where it rapidly forms a viscous colloidal solution with
50 strong activity. It can also incorporate divalent metal ions (excluding mercury and magnesium) to form
51 an alginate gel of high tensile strength within the food.(12-14) Muhammad Lubowa *et al.* prepared rice
52 noodles with pre-gelatinized high amylose corn starch combined with Ca^{2+} induced sodium alginate,
53 which solidified alginate, improved the tensile strength of noodles, and made the noodles more
54 dense.(15) Similarly, Masahiro Yuasa *et al.* used a plastic syringe to add mentsuyu containing 1% (w/w)
55 sodium alginate to 5% (w/w) calcium lactate solution in a drop by drop manner. The solution
56 immediately gelled to form mentsuyu caviar, which gave mentsuyu and caviar an even more striking
57 visual appearance.(16)

58 However, the effect of Ca^{2+} induced sodium alginate gel on the quality and digestive characteristics of
59 buckwheat noodles has not been systematically studied. In this study, two methods of Ca^{2+}
60 incorporation into sodium alginate and its effects on the digestion of pure buckwheat noodles were
61 investigated. The first method was exogenous, where Ca^{2+} permeated sodium alginate-containing
62 noodles from the outside during cooking, resulting in the formation of a stable gel network. The second
63 method was endogenous, where the Ca^{2+} was released from calcium carbonate after acid treatment and
64 subsequent cross-linking with sodium alginate to form the gel network during the procedure of making
65 the noodles. Here, the effects of these different methods of Ca^{2+} incorporation into the sodium alginate
66 networks on the digestion of the noodles were studied, measuring parameters associated with cooking,
67 texture, and *in vitro* digestion of the noodles. In addition, the effects of calcium and sodium alginate
68 cross-linking on the crystal structure of the starch were assessed by X-ray diffraction (XRD) and
69 Fourier transform infrared spectroscopy (FTIR). The results indicate the preferable methods for the
70 manufacture of functional noodles.

71 2 Materials and methods

72 2.1 Materials

73 Pure buckwheat flour was provided by Dalian Hongrun Whole Grain Food Co. *Ltd.* (Dalian, China).
74 Pepsin (≥ 3800 U/g), trypsin (1:4000), and amyloglucosidase (15 U/mL) were purchased from Shanghai
75 Yuanye Bio Biotechnology Co., *Ltd.* (Shanghai, China). Other reagents were obtained from Beijing
76 Chemical Reagent Co. (Beijing, China). All reagents were of analytical grade.

77 2.2 Methods

78 2.2.1 Preparation of noodles

79 Preparation of pure noodles: the noodles were prepared by mixing 90 g of pure buckwheat flour with
80 50 g of deionized water. The optimal cooking time was 3 minutes 40 seconds. It's as the control group.

81 Exogenous method: 90 g of pure buckwheat flour was mixed with 50 g of sodium alginate solution
82 (0.1%-0.5%) using a mixer (HMJ-A35M1, Guangdong, China) for 15 min. After allowing the dough
83 to rest for 15 min, it was sheeted using a noodle machine (MR-08, Guangdong, China) and the sheets
84 were cut into noodles 6.0 mm wide and 2.0 mm thick. Then use the prepared CaCl_2 solution (0.3-0.5M)
85 to cook the noodles until the optimal cooking time: 3 minutes and 40 seconds.

86 Endogenous method: 90 g of pure buckwheat flour was mixed with CaCO_3 powder (3%-9% of the
87 dough) and added to 50 g sodium alginate solution (0.1-0.5%) using a mixer for 15 minutes. After
88 allowing the dough to rest for 15 min, it was sheeted using a noodle machine and the sheets were cut
89 into noodles 6.0 mm wide and 2.0 mm thick. The noodles were cooked with the citric acid solution,
90 pH 4.0, until the optimal cooking time: 3 minutes and 40 seconds.

91 2.2.2 Cooking characteristics of noodles

92 In a separate experiment, the characteristics of the noodles were evaluated according to the method of
93 Gimenez *et al* (17) with slight modifications.

94 Water absorption of noodles: Ten pieces of noodles were weighed on a balance and the weight was
95 recorded as M_1 . The noodles were then boiled in 500 ml of deionized water at 160 °C until cooked.
96 The noodles were then immediately removed from the water, washed with cool water for 10 s, and
97 placed on a screen mesh. After standing at room temperature for 5 min, the noodles were weighed,
98 with the weight recorded as M_2 . The rate of water absorption was calculated using the equation below
99 and the experiment was repeated three times.

$$100 \quad \text{Water absorption(\%)} = \left(\frac{M_2 - M_1}{M_1} \right) \times 100\%$$

101 Turbidity: the cooking water was then cooled and transferred to a 500 ml volume bottle, which was
102 reached volume with deionized water, shaken and left for 2 h, and the absorbance at 460 nm measured
103 by ultraviolet spectrophotometer is turbidity. Measurements were obtained in triplicate.

104 2.2.3 Texture profile analysis

105 The texture profile analysis (TPA) of the noodles was conducted with a texture analyzer (TA-XTC,
106 Boson Tech Co. *Ltd.*, China)(18). The parameters used for the analysis were as follows: P/5 probe, pre-
107 test speed = 2.0 mm/s; test speed = 0.8 mm/s; post-test speed = 0.8 mm/s; compression degree = 50%.
108 Samples were analyzed in octuplicate.

109 2.2.4 Starch digestion *in vitro*

110 The digestibility of the noodles was analyzed *in vitro* as described by Englyst *et al.* with minor
111 modifications. (19) The cooked fresh noodles were ground and the sample (3 g) was mixed with 30
112 mL of distilled water in a beaker and stirred using a magnetic stirrer at 37°C for 10 min. Pepsin (1.0
113 mL) was then added and stirred for 30 min to simulate gastric digestion (pH=2). Once completed, an

114 aliquot (1.0 mL) was withdrawn (time 0) and added to 4 mL absolute alcohol to stop any further enzyme
 115 reaction. Amyloglucosidase (0.1 mL) was then added to the beaker to prevent inhibition of the end
 116 products of pancreatic α -amylase. Then 1 ml 5% trypsin was added to represent ileal digestion.
 117 Aliquots (1.0 mL) were removed at different times (20, 30, 60, 90, 120, and 180 min), which were
 118 inactivated by the addition of absolute ethanol (4.0 mL). Subsequently, the glucose content was
 119 determined by the 3,5-dinitrosalicylic acid (DNS) method.(20) Absorbances at 540 nm were measured
 120 using a UV-3600 spectrophotometer (Shimadzu, Japan) and the amount of hydrolyzed sugar was
 121 calculated as follows:

122

$$123 \quad CHO = C \times D \times (V - S)$$

124 where,

125 CHO: amount of hydrolyzed sugar generated in the whole system during digestion *in vitro* (mg);

126 C: standard amount of glucose detected from the standard operating curve (mg);

127 D: dilution ratio of dialysis solution;

128 V: total volume of the solution for digestion of the whole system *in vitro* (mL);

129 S: volume of solution taken from the system each time (mL).

130 2.2.5 X-ray diffraction analysis

131 The cooked noodles were immediately frozen in a -80 °C freezer. The noodle samples were lyophilized,
 132 ground to powder (SCIENTZ 18N, Zhejiang Side Equipment Co., Ltd, Zhejiang, China) and passed
 133 through a 60-mesh sieve before X-ray diffraction analysis. X-ray diffraction analysis of the noodles
 134 was conducted using an XRD-6000 diffractometer (Shimadzu) with an operating voltage of 40 kV and
 135 a current of 30 mA. The diffraction scan angle (2θ) ranged from 5 to 50° with a scanning speed of
 136 2°/min.

137 2.2.6 FTIR analysis

138 The sample preparation was in a similar manner to the X-ray diffraction analysis. Samples were ground
 139 to a fine powder and were mixed with dry potassium bromide (1:100, w/w) and tableted at 10 000 PSI.
 140 Spectra were recorded in an IR-Affinity-1 spectrophotometer (Shimadzu) between 399 and 4000 cm^{-1}
 141 at a resolution of 2 cm^{-1} .

142 2.2.7 Statistical analysis

143 All experiments were performed with three replicates. Data were expressed as mean \pm standard
 144 deviation and analyzed using Origin 2021 (Origin-Lab, Inc., USA) and GraphPad Prism 8.4.0
 145 (GraphPad Software, LLC, USA) for Windows. Differences between means were assessed by one-way
 146 analysis of variance (ANOVA) followed by Duncan's test.

147 3 Results and Discussion

148 3.1 Cooking characteristics of noodles

149 The results of the water absorption and turbidity of noodles prepared by the exogenous method are
 150 shown in Table 1. Water absorption is defined as the ability of the noodles to retain water, and is
 151 controlled mainly by the strength of the network formed by starch, fiber, or protein.(21) Compared
 152 with the control groups in both exogenous and endogenous methods, the water absorption of noodles
 153 supplemented with sodium alginate colloid was significantly reduced, and further decreases were seen
 154 in the noodles in which calcium ions had been incorporated with the sodium alginate ($p < 0.05$). This

155 may be because the capacity for water absorption is related to the integrity of the structural network,
 156 and a highly cross-linked network structure limits water absorption.(22) The noodles cooked by the
 157 exogenous method are more turbid than the noodle soup of the control group. The possible reason is
 158 that during the cooking process, part of calcium chloride entered the noodles and combined with
 159 sodium alginate to form a network structure, while the other part of calcium chloride remained in the
 160 noodle soup, increasing the turbidity degree of the noodle soup. However, different from the exogenous
 161 method, the turbidity of noodles prepared by the endogenous method decreased with the increase of
 162 the concentrations of sodium alginate and calcium carbonate, which may be caused by the increase of
 163 the concentration of both, which further promoted the formation of gel network structure.

164 3.2 Textural properties of noodles

165 Fig. 1 illustrates the textural properties of the noodles. It was found that, compared with the control
 166 noodles, the hardness, springiness, cohesiveness, adhesiveness, chewiness, and other textural
 167 properties were enhanced by the addition of sodium alginate at different concentrations (Fig.1A). The
 168 hardness, adhesiveness, and chewiness of noodles with Ca^{2+} incorporated into the sodium alginate by
 169 the exogenous method were significantly higher than those in the sodium alginate only groups (Fig.1B-
 170 D). The increase in these properties may be due to the effective cross-linking of Ca^{2+} with sodium
 171 alginate, resulting in the strengthening of the three-dimensional structures. However, compared with
 172 the exogenous method, there were no significant differences in the textural characteristics between the
 173 noodles prepared by the endogenous method with Ca^{2+} and the sodium alginate only groups (Fig.2A-
 174 D), which may be due to the fact that citric acid is a weak electrolyte with weak ionizing ability, and
 175 the reaction with calcium carbonate releases less calcium ions, resulting in a poor ability to bind sodium
 176 alginate and not form a more stable strong gel. These results confirm that the gel network structure of
 177 Ca^{2+} and sodium alginate formed by the exogenous method could improve the texture of noodles.

178 3.3 Digestibility of noodles *in vitro*

179 Fig 2 shows the amounts of reduced sugars released over 180 min of *in vitro* digestion for all the noodle
 180 samples. The same trend of starch hydrolysis was seen in all samples, with the amount of glucose
 181 released increasing gradually over time. Compared with the control noodles, the addition of different
 182 concentrations of sodium alginate (0.1%-0.5%) did not reduce the glucose release. (Fig.3A) However,
 183 the incorporation of Ca^{2+} into the sodium alginate-containing noodles using the exogenous method
 184 resulted in reductions in glucose release, with noodles containing higher concentrations of Ca^{2+}
 185 showing lower glucose release; the maximum reduction in glucose release was approximately 23.3
 186 mg/g.(Fig.3B-D) This is likely the consequence of the denser network formed by the calcium and
 187 sodium alginate, preventing the leaching of internal molecules. These findings indicate that the network
 188 composed of sodium alginate and calcium ions can protect the starch from amylase hydrolysis. The
 189 concomitant use of higher concentrations of Ca^{2+} results in a denser gel network that provides a better
 190 barrier to enzymatic hydrolysis. These results are consistent with previous findings where Ca^{2+} was
 191 used for cross-linking with sodium alginate to form a core-shell structured macrocapsule calcium
 192 alginate.(23)

193 The glucose release of noodles prepared by the endogenous method of calcium ion combined with
 194 sodium alginate is shown in Fig.4.(A-D). The glucose release was lower than that in the sodium
 195 alginate group but was not decreased in comparison with the control group. Although increased Ca^{2+}
 196 concentrations were beneficial to glucose release compared with the sodium alginate only groups, the
 197 incorporation of calcium did not significantly reduce the glucose release compared with the control
 198 group. The likely explanation for this phenomenon may be as below. On the one hand, the relatively
 199 low concentrations of calcium ions were released by the reaction of calcium carbonate with acid,

200 resulting in the formation of a weaker gel structure. On the other hand, the carbon dioxide formed
201 during the reaction may affect the structure of the dough, creating greater looseness, which may also
202 account for the poorer textural properties.

203

204 3.4 XRD patterns of starches

205 XRD was used to investigate changes in the crystallinity of noodles prepared by the exogenous method.
206 The XRD patterns of all the samples are summarized in Fig. 5. The crystallinity of foods is a significant
207 determinant of their physical properties, and affects digestibility.(24) Starch is composed of four types
208 of crystal structures, namely, A, B, C, and V.(25) As seen in the XRD patterns in the figure, clear peaks
209 are visible at 2θ of 20° , which indicate the possible presence of an amylose-lipid complex (V-type) in
210 the starch particles and endows the starch with properties such as resistance to digestion and improved
211 food texture.(26) The XRD patterns of the noodles with sodium alginate and calcium were similar to
212 those of the sodium alginate only noodles; however, the area of the peak was altered by the addition of
213 calcium ions. The X-ray diffraction pattern results of the endogenous method are similar to those of
214 the exogenous method, which only changed the crystal area of the noodles, which are not mentioned
215 here. This indicates that the crystal structure of the starch was not changed by the cross-linking of
216 calcium ions with sodium alginate, with the addition of calcium altering only the crystal area of the
217 noodles, possibly affecting the quality of the noodles.(27)

218 3.5 FTIR spectroscopy of starches

219 The FTIR spectra in the wavenumber range of $399\text{--}4000\text{ cm}^{-1}$ of starches with exogenously
220 incorporated Ca^{2+} and sodium alginate are shown in Fig. 6. As seen in the figure, the spectra of the
221 different samples are similar. The absorption peak at 1652 cm^{-1} results from the stretching vibration of
222 $\text{C}=\text{O}$, also seen in α -helical structures of proteins.(28) The absorption peak at 2927 cm^{-1} results from
223 the asymmetric stretching vibration of CH_2 .(29) Major absorption peaks are visible in the hydroxyl
224 region (centered at $3394\text{--}3423\text{ cm}^{-1}$), likely the result of hydrogen bonds between the starch particles,
225 alginate, and gel polysaccharide molecules.(30) Compared with the sodium alginate only samples, the
226 addition of calcium ions did not result in a new absorption peak, indicating that no chemical bonds
227 were formed between sodium alginate and calcium in the noodles. The Fourier infrared spectrogram
228 results of the endogenous method are similar to those of the exogenous method, with no significant
229 changes, which are not mentioned here.

230 4 Conclusions

231 We prepared novel types of functional noodles by two simple methods and studied their properties in
232 relation to cooking, texture, and digestion, as well as analyzing the structure of the starch. It was found
233 that the stable cross-linking system formed by Ca^{2+} and sodium alginate could reduce the rate of water
234 absorption by the noodles, and improve the textural properties of the noodles, such as hardness,
235 springiness, cohesiveness, adhesiveness, and chewiness. In addition, the formation of the alginate
236 network structure altered the area of the crystal zone of the noodles, although no new chemical bonds
237 or substances were generated. Most importantly, the cross-linking of Ca^{2+} and sodium alginate
238 significantly reduced the amount of glucose released from the noodles. In conclusion, the noodles
239 prepared by the exogenous method were superior to those prepared using the endogenous method in
240 terms of both noodle quality and lower glucose release, which will contribute to the development of
241 functional foods. However, these results were obtained using simulated digestion *in vitro*, and further
242 *in vivo* investigations are needed for verification.

243 **5 Conflict of Interest**

244 The authors declare that the research was conducted in the absence of any commercial or financial
245 relationships that could be construed as a potential conflict of interest.

246 **6 Author Contributions**

247 **Bing Hu, Jixin Yang** and **Jijuan Cao** contributed to the conception of the study;
248 **Hongyan Wang** performed the experiment;
249 **Jiukai Zhang** and **Lingyu Han** contributed significantly to analysis and manuscript preparation;
250 **Hongyan Wang** performed the data analyses and wrote the manuscript;
251 **Ying Zhang** helped perform the analysis with constructive discussions.

252 **7 Funding**

253 This article was funded by the National Key Research and Development Project (2021YFF0601900),
254 Liaoning Provincial Science and Technology Innovation Leading Talents Project (XLYC2002106),
255 Natural Science Foundation of Liaoning Province (2021-MS-147), Dalian High-Level Talent
256 Innovation, Scientific and Technological Talent Entrepreneurship and Innovation Team Support
257 Projects in Key Fields (2020RQ122), Liaoning Province Livelihood Science and Technology Project
258 (2021JH2/10200019), Department of Education of Liaoning Province (LJKZ0037), Dalian Key
259 Science And Technology Project (2021JB12SN038).

260 **8 Reference styles**

261 [1] World Health Organization. (2016). Global report on diabetes. World Health Organization.
262 <https://apps.who.int/iris/handle/10665/204871>.

263 [2] Liu, D.; Song, S.; Tao, L.; Yu, L.; Wang, J., Effects of common buckwheat bran on wheat dough
264 properties and noodle quality compared with common buckwheat hull. *Lwt* 2022,155.
265 doi.org/10.1016/j.lwt.2021.112971

266 [3] Gan, C.Y.; Ong, W.H.; Wong, L.M.; Easa, A. M., Effects of ribose, microbial transglutaminase and
267 soy protein isolate on physical properties and in-vitro starch digestibility of yellow noodles. *LWT -*
268 *Food Science and Technology* 2009, 42 (1), 174-179. [doi:10.1016/j.lwt.2008.05.004](https://doi.org/10.1016/j.lwt.2008.05.004)

269 [4] Atkinson, F. S.; Foster-Powell, K.; Brand-Miller, J. C., International tables of glycemic index and
270 glycemic load values: 2008. *Diabetes Care* 2008, 31 (12), 2281-3. [doi:10.2337/dc08-1239](https://doi.org/10.2337/dc08-1239)

271 [5] Zhu, F., Chemical composition and health effects of Tartary buckwheat. *Food Chemistry* 2016,
272 203, 231-245. [doi:10.1016/j.foodchem.2016.02.050](https://doi.org/10.1016/j.foodchem.2016.02.050).

273 [6] Schoenlechner R, Drausinger J, Ottenschlaeger V, Jurackova K, Berghofer E Functional properties
274 of gluten-free pasta produced from amaranth, quinoa and buckwheat. *Plant Foods Hum Nutr* 2010,
275 65:339–349. [doi:10.1007/s11130-010-0194-0](https://doi.org/10.1007/s11130-010-0194-0)

276 [7] Jia, B.; Yao, Y.; Liu, J.; Guan, W.; Brennan, C. S.; Brennan, M. A., Physical Properties and In Vitro
277 Starch Digestibility of Noodles Substituted with Tartary Buckwheat Flour. *Starch - Stärke* 2019.
278 [doi:10.1002/star.201800314](https://doi.org/10.1002/star.201800314)

- 279 [8] Lucas-Gonzalez, R.; Angel Perez-Alvarez, J.; Moscaritolo, S.; Fernandez-Lopez, J.; Sacchetti, G.;
 280 Viuda-Martos, M., Evaluation of polyphenol bioaccessibility and kinetic of starch digestion of
 281 spaghetti with persimmon (*Diospyros kaki*) flours coproducts during in vitro gastrointestinal digestion.
 282 *Food Chemistry* 2021, 338, 128142. doi:10.1016/j.foodchem.2020.128142
- 283 [9] Wang, C.; Li, W.; Chen, Z.; Gao, X.; Yuan, G.; Pan, Y.; Chen, H., Effects of simulated
 284 gastrointestinal digestion in vitro on the chemical properties, antioxidant activity, alpha-amylase and
 285 alpha-glucosidase inhibitory activity of polysaccharides from *Inonotus obliquus*. *Food Research*
 286 *International* 2018, 103, 280-288. doi:10.1016/j.foodres.2017.10.058
- 287 [10] Foschia, M.; Peressini, D.; Sensidoni, A.; Brennan, M. A.; Brennan, C. S., Synergistic effect of
 288 different dietary fibres in pasta on in vitro starch digestion? *Food Chemistry* 2015, 172, 245-50.
 289 doi:10.1016/j.foodchem.2014.09.062 doi:10.1016/j.lwt.2020.110398
- 290 [11] Lubowa, M.; Yeoh, S. Y.; Varastegan, B.; Easa, A. M., Effect of pre-gelatinised high-amylose
 291 maize starch combined with Ca²⁺ induced setting of alginate on the physicochemical and sensory
 292 properties of rice flour noodles. *International Journal of Food Science & Technology* 2020, 56 (2),
 293 1021-1029. doi:10.1111/ijfs.14754
- 294 [12] Hong, T.; Zhang, Y.; Xu, D.; Wu, F.; Xu, X., Effect of sodium alginate on the quality of highland
 295 barley fortified wheat noodles. *Lwt* 2021, 140. doi:10.1016/j.lwt.2020.110719
- 296 [13] Kakita, H.; Kamishima, H., Some properties of alginate gels derived from algal sodium alginate.
 297 *Journal of Applied Phycology* 2008, 20 (5), 543-549. doi:10.1007/s10811-008-9317-5
- 298 [14] Koh, L. W.; Kasapis, S.; Lim, K. M.; Foo, C. W., Structural enhancement leading to retardation of
 299 in vitro digestion of rice dough in the presence of alginate. *Food Hydrocolloids* 2009, 23 (6), 1458-
 300 1464. doi:10.1016/j.foodhyd.2008.07.020
- 301 [15] Muhammad, L., Yeoh, S., Varastegan, B., & Easa, A. M. Effect of pre-gelatinized high amylose
 302 maize starch combined with Ca²⁺ -induced setting of alginate on the physicochemical and sensory
 303 properties of rice flour noodles. *International Journal of Food Science &*
 304 *Technology*. doi:10.1111/ijfs.14754
- 305 [16] Yuasa, M., Tagawa, Y., & Tominaga, M. . The texture and preference of “mentsuyu (Japanese
 306 noodle soup base) caviar” prepared from sodium alginate and calcium lactate. *International Journal of*
 307 *Gastronomy and Food Science*, 18, 100178. doi:10.1016/j.ijgfs.2019.100178
- 308 [17] Gimenez, M. A.; Gonzalez, R. J.; Wagner, J.; Torres, R.; Lobo, M. O.; Samman, N. C., Effect of
 309 extrusion conditions on physicochemical and sensorial properties of corn-broad beans (*Vicia faba*)
 310 spaghetti type pasta. *Food Chemistry* 2013, 136 (2), 538-545. doi:10.1016/j.foodchem.2012.08.068
- 311 [18] Deng, Y.; Wu, Y.; Li, Y., Physiological responses and quality attributes of ‘Kyoho’grapes to
 312 controlled atmosphere storage. *LWT- Food Science and Technology* 2006, 39 (6), 584-590.
 313 doi:10.1016/j.lwt.2005.05.001
- 314 [19] Englyst, H. N.; Kingman, S. M.; Cummings, J., Classification and measurement of nutritionally
 315 important starch fractions. *European journal of clinical nutrition* 1992, 46, S33-50.

- 316 [20] Brennan, M. A., Derbyshire, E., Tiwari, B. K., & Brennan, C. S.. Ready-to-eat snack products: the
317 role of extrusion technology in developing consumer acceptable and nutritious snacks. *International*
318 *Journal of Food Science & Technology*, 2013, 48(5), 893–902. doi:10.1111/ijfs.12055
- 319 [21] Gupta, A., Sharma, S., & Reddy Surasani, V. K.. Quinoa protein isolate supplemented pasta:
320 Nutritional, physical, textural and morphological characterization. *LWT*,
321 2020,110045. doi:10.1016/j.lwt.2020.110045
- 322 [22] Sandhu, K. S.; Kaur, M.; Mukesh, Studies on noodle quality of potato and rice starches and their
323 blends in relation to their physicochemical, pasting and gel textural properties. *LWT - Food Science*
324 *and Technology* 2010, 43 (8), 1289-1293. doi:10.1016/j.lwt.2010.03.003
- 325 [23] Cui, C.; Li, M.; Ji, N.; Qin, Y.; Shi, R.; Qiao, Y.; Xiong, L.; Dai, L.; Sun, Q.. Calcium
326 alginate/curdlan/corn starch@calcium alginate macrocapsules for slowly digestible and resistant starch.
327 *Carbohydr Polym* 2022, 285, 119259. doi:10.1016/j.carbpol.2022.119259
- 328 [24] Dona, A. C.; Pages, G.; Gilbert, R. G.; Kuchel, P. W.. Digestion of starch: In vivo and in vitro
329 kinetic models used to characterise oligosaccharide or glucose release. *Carbohydrate Polymers* 2010,
330 80 (3), 599-617. doi:10.1016/j.carbpol.2010.01.002
- 331 [25] Liu, Y.; Xu, Y.; Yan, Y.; Hu, D.; Yang, L.; Shen, R.. Application of Raman spectroscopy in
332 structure analysis and crystallinity calculation of corn starch. *Starch - Stärke* 2015, 67 (7-8), 612-619.
333 doi:10.1002/star.201400246
- 334 [26] Hung, P. V.; Vien, N. L.; Lan Phi, N. T.. Resistant starch improvement of rice starches under a
335 combination of acid and heat-moisture treatments. *Food Chemistry* 2016, 191, 67-73.
336 doi:10.1016/j.foodchem.2015.02.002
- 337 [27] Jia, M.; Yu, Q.; Chen, J.; He, Z.; Chen, Y.; Xie, J.; Nie, S.; Xie, M.. Physical quality and in vitro
338 starch digestibility of biscuits as affected by addition of soluble dietary fiber from defatted rice bran.
339 *Food Hydrocolloids* 2020, 99.10. doi:1016/j.foodhyd.2019.105349
- 340 [28] Jeong, S.; Kim, H. W.; Lee, S.. Rheological and secondary structural characterization of rice flour-
341 zein composites for noodles slit from gluten-free sheeted dough. *Food Chem* 2017, 221, 1539-1545.
342 doi:10.1016/j.foodchem.2016.10.139
- 343 [29] Flores-Morales, A.; Jimenez-Estrada, M.; Mora-Escobedo, R.. Determination of the structural
344 changes by FT-IR, Raman, and CP/MAS (13)C NMR spectroscopy on retrograded starch of maize
345 tortillas. *Carbohydr Polym* 2012, 87 (1), 61-68. doi: 10.1016/j.carbpol.2011.07.011
- 346 [30] Li, Q.; Duan, M.; Hou, D.; Chen, X.; Shi, J.; Zhou, W.. Fabrication and characterization of
347 Ca(II)-alginate-based beads combined with different polysaccharides as vehicles for delivery, release
348 and storage of tea polyphenols. *Food Hydrocolloids* 2021,112,106274. doi:112.10.1016/j.foodhyd

349 Table1. Cooking characteristics of noodle samples

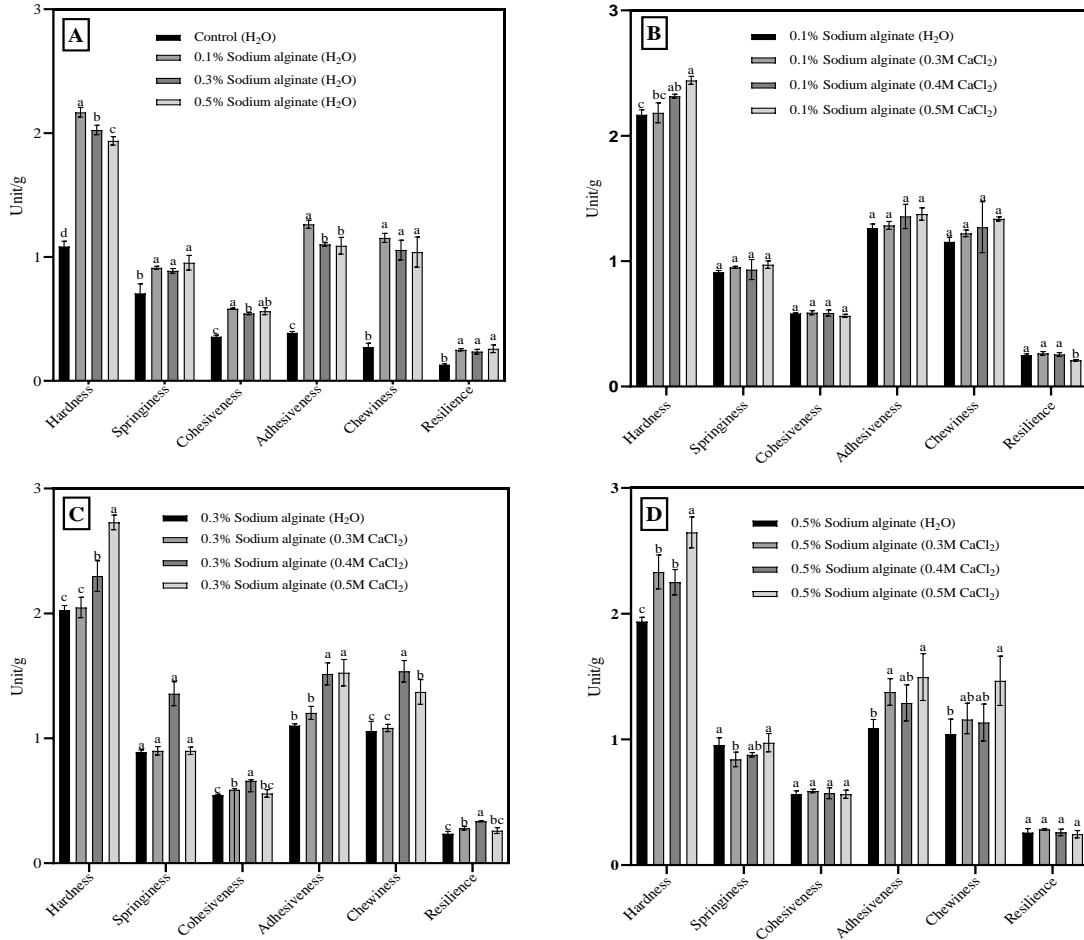
Method	Noodle Samples	Water Absorption(%)	Turbidity
Exogenous method	Control (H ₂ O)*	131.727±3.285 ^a	0.069±0.002 ^g
	0.1% Sodium alginate (H ₂ O)*	119.330±5.064 ^b	0.092±0.031 ^{fg}
	0.1% Sodium alginate (0.3M CaCl ₂)*	109.287±5.500 ^{de}	0.156±0.002 ^{bc}
	0.1% Sodium alginate (0.4M CaCl ₂)*	109.537±2.577 ^{cde}	0.208±0.001 ^a
	0.1% Sodium alginate (0.5M CaCl ₂)*	102.009±0.299 ^e	0.160±0.002 ^{bc}
	0.3% Sodium alginate (H ₂ O)*	116.808±14.879 ^{bc}	0.109±0.001 ^{ef}
	0.3% Sodium alginate (0.3M CaCl ₂)*	112.768±1.935 ^{bcd}	0.148±0.003 ^{bcd}
	0.3% Sodium alginate (0.4M CaCl ₂)*	111.246±0.857 ^{bcd}	0.131±0.007 ^{cde}
	0.3% Sodium alginate (0.5M CaCl ₂)*	106.494±1.783 ^{de}	0.163±0.007 ^{bc}
	0.5% Sodium alginate (H ₂ O)*	117.858±1.224 ^{bc}	0.101±0.002 ^g
	0.5% Sodium alginate (0.3M CaCl ₂)*	116.906±4.045 ^{bc}	0.118±0.014 ^{def}
	0.5% Sodium alginate (0.4M CaCl ₂)*	108.145±2.916 ^{cde}	0.112±0.001 ^{ef}
	0.5% Sodium alginate (0.5M CaCl ₂)*	105.210±2.806 ^{de}	0.174±0.004 ^b
Endogenous method	Control (Citric acid)*	133.759±0.858 ^a	0.074±0.003 ^g
	0.1% Sodium alginate (Citric acid)*	133.501±3.289 ^a	0.147±0.004 ^b
	0.1% Sodium alginate/3% CaCO ₃ (Citric acid)*	129.970±5.575 ^{ab}	0.143±0.005 ^b
	0.1% Sodium alginate/6% CaCO ₃ (Citric acid)*	123.784±1.967 ^{bcd}	0.140±0.003 ^b
	0.1% Sodium alginate/9% CaCO ₃ (Citric acid)*	117.172±1.497 ^{de}	0.119±0.007 ^{cd}
	0.3% Sodium alginate (Citric acid)*	123.001±0.055 ^{bcd}	0.123±0.006 ^c
	0.3% Sodium alginate/3% CaCO ₃ (Citric acid)*	119.832±9.728 ^{cde}	0.118±0.003 ^{cd}
	0.3% Sodium alginate/6% CaCO ₃ (Citric acid)*	117.986±8.109 ^{de}	0.118±0.004 ^{cd}
	0.3% Sodium alginate/9%CaCO ₃ (Citric acid)*	115.233±2.219 ^e	0.115±0.002 ^{cd}
	0.5% Sodium alginate (Citric acid)*	132.062±1.120 ^a	0.156±0.004 ^a
	0.5% Sodium alginate/3%CaCO ₃ (Citric acid)*	127.251±2.171 ^{abc}	0.113±0.005 ^d
	0.5% Sodium alginate/6% CaCO ₃ (Citric acid)*	121.569±1.825 ^{cde}	0.105±0.002 ^e
	0.5% Sodium alginate/9% CaCO ₃ (Citric acid)*	114.959±3.357 ^e	0.087±0.003 ^f

350

351 *Represents different solution environments for cooking noodles.

352 Results are presented as means ± standard deviations. Lowercase letters within columns represent
 353 significant differences (P < 0.05) between samples.

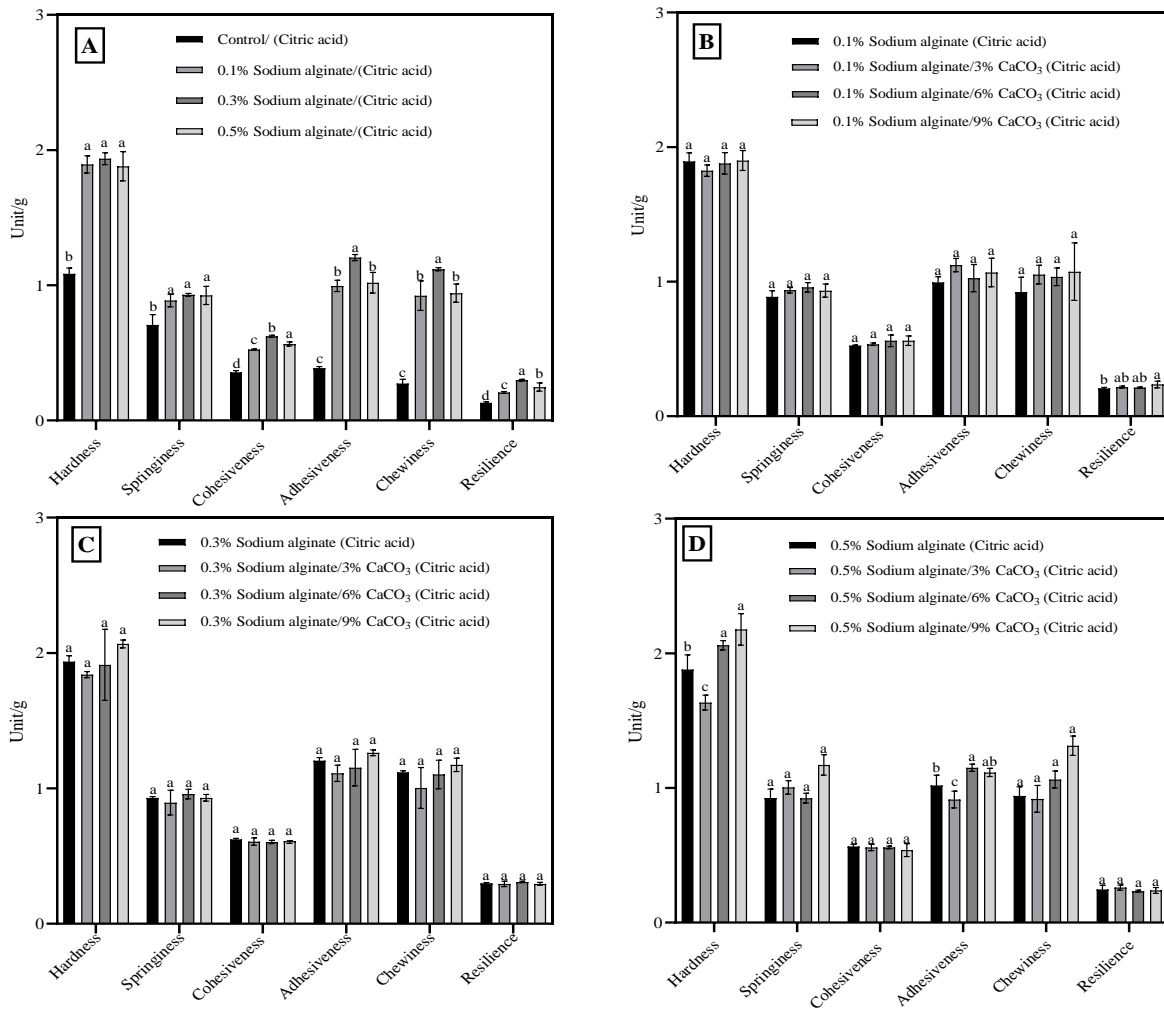
354



355

356 Fig. 1. A-D show the effects of varying Ca²⁺ concentrations on the textural properties of buckwheat
 357 noodles prepared by the exogenous method. Control group (A) was steamed with deionized water. The
 358 experimental group in B, C and D were steamed with calcium chloride solution of different
 359 concentrations.

360



361 Fig. 2. A-D show the effects of Ca^{2+} concentrations of the textural properties of noodles prepared by
 362 the endogenous method. The solution environment for cooking noodles in A-D is 0.1g/L citric acid
 363 solution (pH=4).

364

365

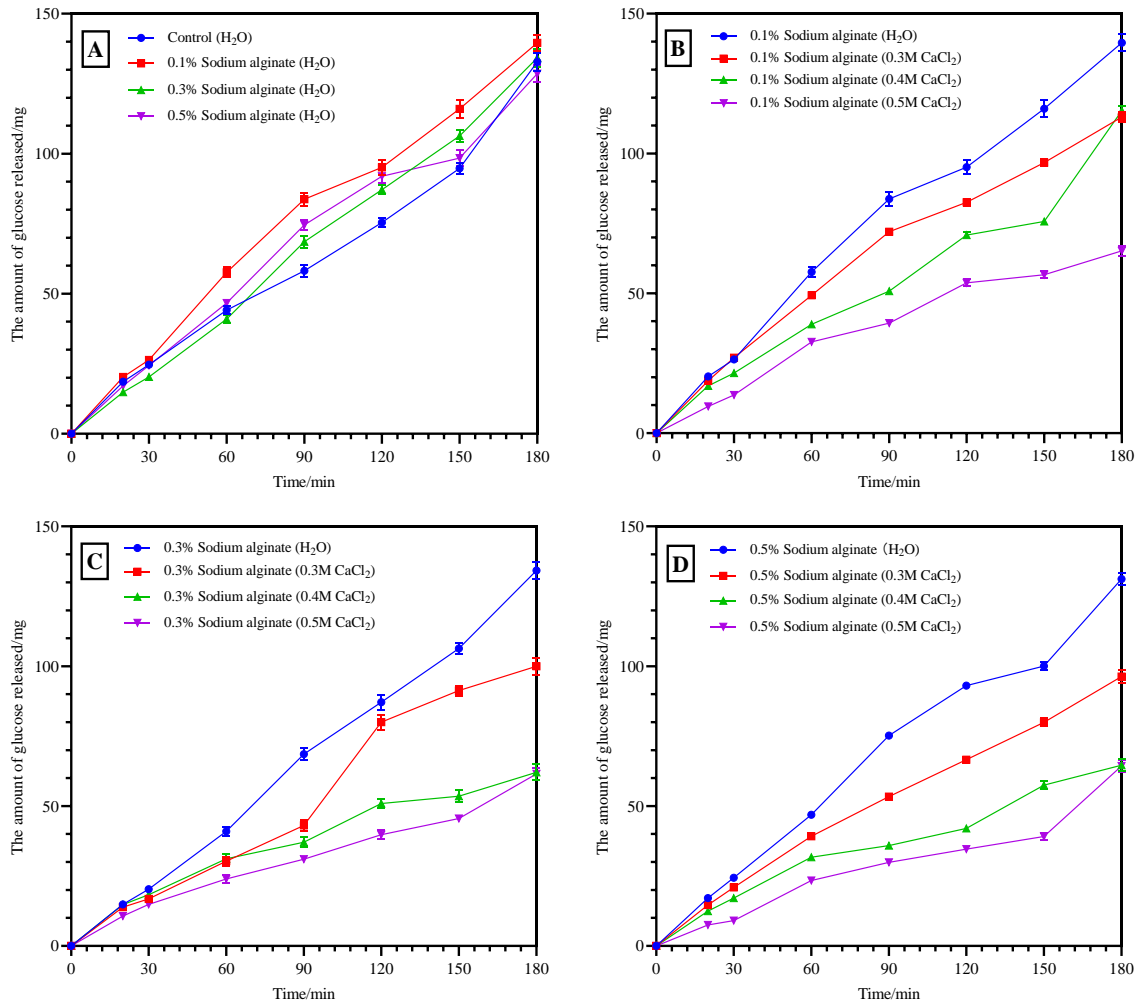
366

367

368

369

370

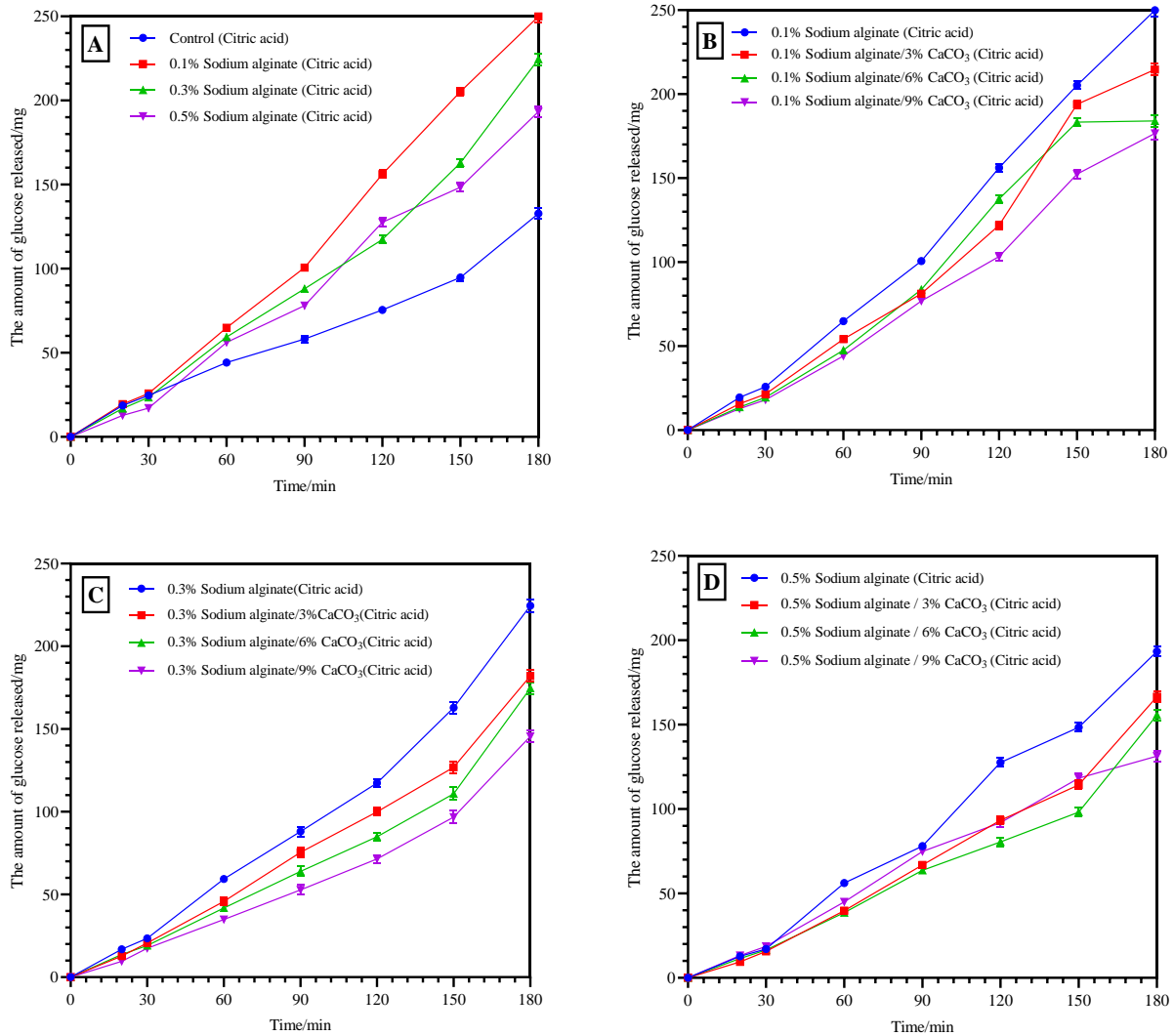


371

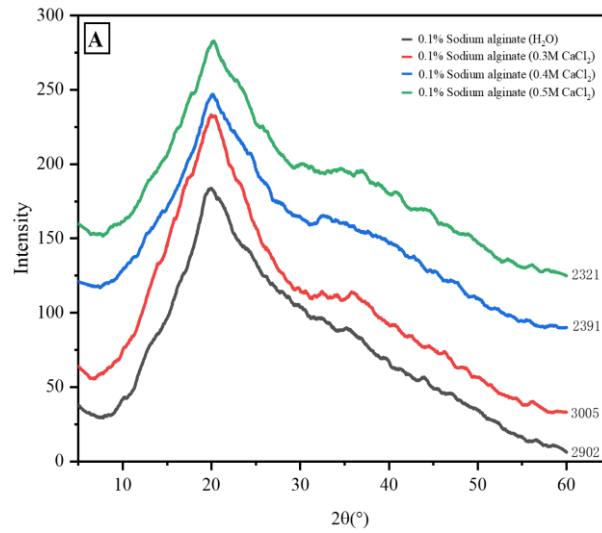
372 Fig. 3. A-D show the effects of varying Ca²⁺ concentrations of the digestibility of buckwheat noodles
 373 prepared by the exogenous method. Control group (A) was steamed with deionized water. The
 374 experimental groups (B, C, D) were steamed with calcium chloride solution of different concentrations.

375

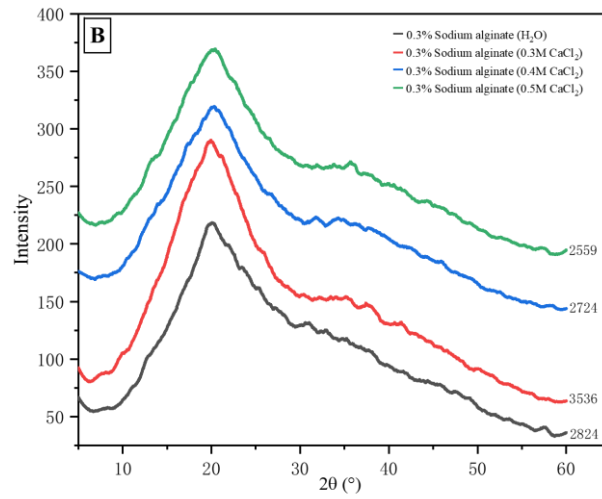
376



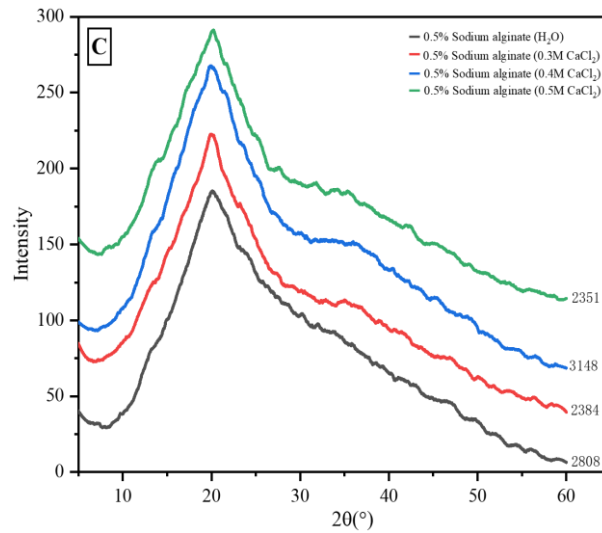
377
 378 Fig. 4. A-D show the effects of varying Ca²⁺ concentrations on the digestibility of buckwheat noodles
 379 prepared by the endogenous method. The solution environment for cooking noodles in A-D is 0.2 g/L
 380 citric acid solution (pH=4).



381

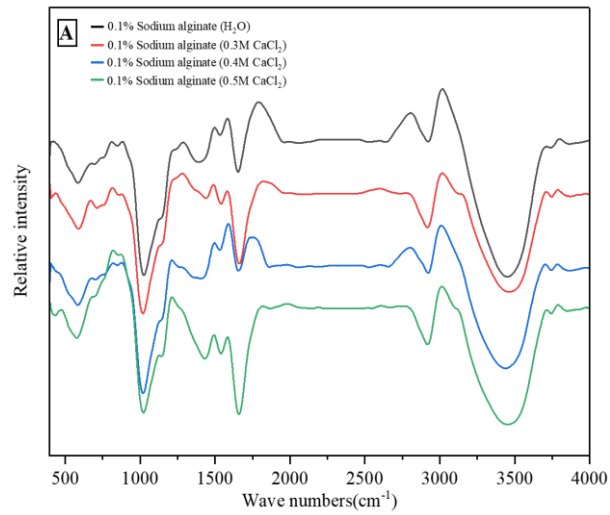


382

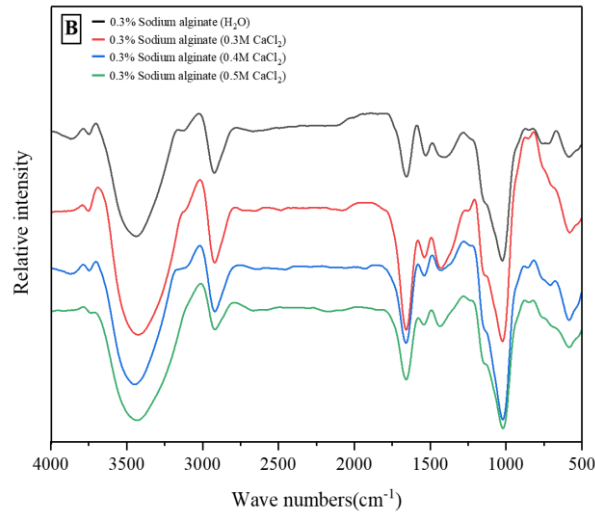


383

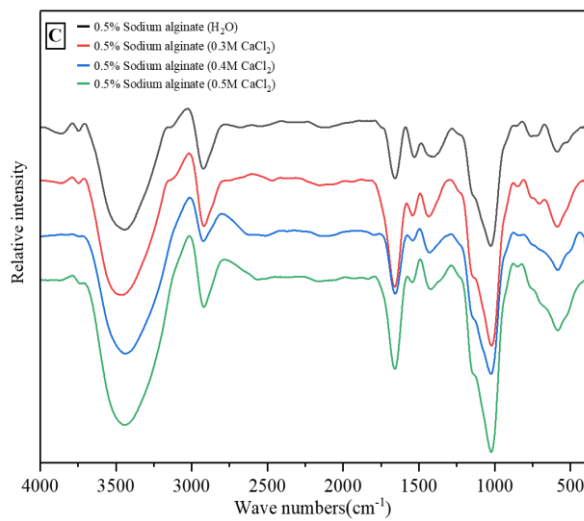
384 Fig. 5. A-C show the X-ray diffraction patterns of sodium alginate with the addition of varying Ca^{2+}
 385 concentrations by the exogenous method.



386



387



388

389 Fig. 6. A-C show the Fourier transform infrared spectra of sodium alginate with the addition of varying
 390 Ca^{2+} concentrations by the exogenous method.